

# UNCLASSIFIED

AD NUMBER
ADB262544
NEW LIMITATION CHANGE
TO Approved for public release, distribution unlimited
FROM Distribution authorized to U.S. Gov't. agencies only; Proprietary Info.; Oct 99. Other requests shall be referred to U.S. Army Medical Research and Materiel Command, 504 Scott St., Fort Detrick, MD 21702-5012.
AUTHORITY
USAMRMC- Ft Detrick MD, ltr dtd 24 Jul 2001.

THIS PAGE IS UNCLASSIFIED

AD \_\_\_\_\_

Award Number: DAMD17-97-1-7114

TITLE: Bone Factors Regulating the Osteotropism of Metastatic  
Breast Cancer

PRINCIPAL INVESTIGATOR: Peter V. Hauschka, Ph.D.

CONTRACTING ORGANIZATION: Children's Hospital Corporation  
Boston, Massachusetts 02115

REPORT DATE: October 1999

TYPE OF REPORT: Annual

PREPARED FOR: U.S. Army Medical Research and Materiel Command  
Fort Detrick, Maryland 21702-5012

DISTRIBUTION STATEMENT: Distribution authorized to U.S. Government  
agencies only (proprietary information, Oct 99). Other requests  
for this document shall be referred to U.S. Army Medical Research  
and Materiel Command, 504 Scott Street, Fort Detrick, Maryland  
21702-5012.

The views, opinions and/or findings contained in this report are  
those of the author(s) and should not be construed as an official  
Department of the Army position, policy or decision unless so  
designated by other documentation.

DTIC QUALITY INSPECTED 4

20010122 127

## NOTICE

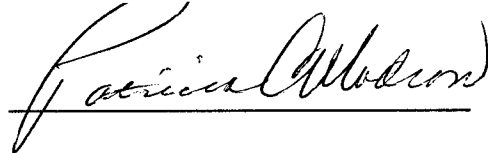
USING GOVERNMENT DRAWINGS, SPECIFICATIONS, OR OTHER DATA INCLUDED IN THIS DOCUMENT FOR ANY PURPOSE OTHER THAN GOVERNMENT PROCUREMENT DOES NOT IN ANY WAY OBLIGATE THE U.S. GOVERNMENT. THE FACT THAT THE GOVERNMENT FORMULATED OR SUPPLIED THE DRAWINGS, SPECIFICATIONS, OR OTHER DATA DOES NOT LICENSE THE HOLDER OR ANY OTHER PERSON OR CORPORATION; OR CONVEY ANY RIGHTS OR PERMISSION TO MANUFACTURE, USE, OR SELL ANY PATENTED INVENTION THAT MAY RELATE TO THEM.

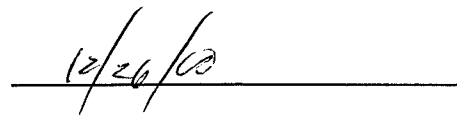
### LIMITED RIGHTS LEGEND

Award Number: DAMD17-97-1-7114  
Organization: Children's Hospital Corporation  
Location of Limited Rights Data (Pages):

Those portions of the technical data contained in this report marked as limited rights data shall not, without the written permission of the above contractor, be (a) released or disclosed outside the government, (b) used by the Government for manufacture or, in the case of computer software documentation, for preparing the same or similar computer software, or (c) used by a party other than the Government, except that the Government may release or disclose technical data to persons outside the Government, or permit the use of technical data by such persons, if (i) such release, disclosure, or use is necessary for emergency repair or overhaul or (ii) is a release or disclosure of technical data (other than detailed manufacturing or process data) to, or use of such data by, a foreign government that is in the interest of the Government and is required for evaluational or informational purposes, provided in either case that such release, disclosure or use is made subject to a prohibition that the person to whom the data is released or disclosed may not further use, release or disclose such data, and the contractor or subcontractor or subcontractor asserting the restriction is notified of such release, disclosure or use. This legend, together with the indications of the portions of this data which are subject to such limitations, shall be included on any reproduction hereof which includes any part of the portions subject to such limitations.

THIS TECHNICAL REPORT HAS BEEN REVIEWED AND IS APPROVED FOR PUBLICATION.

  
\_\_\_\_\_

  
\_\_\_\_\_

**REPORT DOCUMENTATION PAGE**Form Approved  
OMB No. 074-0188

Public reporting burden for this collection of information is estimated to average 1 hour per response, including the time for reviewing instructions, searching existing data sources, gathering and maintaining the data needed, and completing and reviewing this collection of information. Send comments regarding this burden estimate or any other aspect of this collection of information, including suggestions for reducing this burden to Washington Headquarters Services, Directorate for Information Operations and Reports, 1215 Jefferson Davis Highway, Suite 1204, Arlington, VA 22202-4302, and to the Office of Management and Budget, Paperwork Reduction Project (0704-0188), Washington, DC 20503

<b>1. AGENCY USE ONLY (Leave blank)</b>		<b>2. REPORT DATE</b> October 1999	<b>3. REPORT TYPE AND DATES COVERED</b> Annual (22 Sep 98 - 21 Sep 99)	
<b>4. TITLE AND SUBTITLE</b> Bone Factors Regulating the Osteotropism of Metastatic Breast Cancer			<b>5. FUNDING NUMBERS</b> DAMD17-97-1-7114	
<b>6. AUTHOR(S)</b> Peter V. Hauschka, Ph.D.				
<b>7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES)</b> Children's Hospital Corporation Boston, Massachusetts 02115  E-MAIL: hauschka@hub.tch.harvard.edu			<b>8. PERFORMING ORGANIZATION REPORT NUMBER</b>	
<b>9. SPONSORING / MONITORING AGENCY NAME(S) AND ADDRESS(ES)</b> U.S. Army Medical Research and Materiel Command Fort Detrick, Maryland 21702-5012			<b>10. SPONSORING / MONITORING AGENCY REPORT NUMBER</b>	
<b>11. SUPPLEMENTARY NOTES</b>				
<b>12a. DISTRIBUTION STATEMENT:</b> Distribution authorized to U.S. Government agencies only (proprietary information, Oct 99). Other requests for this document shall be referred to U.S. Army Medical Research and Materiel Command, 504 Scott Street, Fort Detrick, Maryland 21702-5012.				<b>12b. DISTRIBUTION CODE</b>
<b>13. ABSTRACT (Maximum 200 Words)</b>  The specificity with which breast cancer metastasizes to the human skeleton is currently unexplained. In this project we have attempted to identify the critical molecular interactions and cellular processes responsible for this osteotropism (homing and metastatic growth in bone) of breast cancer. The central hypothesis is that malignant breast adenocarcinoma cells subvert the cooperative paracrine interactions between normal bone cells, endothelial cells, and the extracellular matrix in order to establish metastatic foci. Our data combined with published reports show that invasive breast adenocarcinoma cells mimic certain hallmarks of the osteoblast phenotype, possibly explaining the facile growth and survival of these tumor cells in bone. Analysis of model human tumor lines, and derivative sublines recovered from bone metastases in nude mice, reveals a correlation between the degree of malignancy and the inappropriate expression of the "bone specific" transcription factor CBFA1. The osteoblast mimicry by the tumor cells may be explained by CBFA1 expression; normal breast tissue does not express CBFA1. Osteoblast factors also have a paracrine effect on altering the phenotype of breast adenocarcinoma cells in the bone microenvironment.				
<b>14. SUBJECT TERMS</b> Breast Cancer			<b>15. NUMBER OF PAGES</b> 33	
			<b>16. PRICE CODE</b>	
<b>17. SECURITY CLASSIFICATION OF REPORT</b> Unclassified	<b>18. SECURITY CLASSIFICATION OF THIS PAGE</b> Unclassified	<b>19. SECURITY CLASSIFICATION OF ABSTRACT</b> Unclassified	<b>20. LIMITATION OF ABSTRACT</b> Limited	

NSN 7540-01-280-5500

Standard Form 298 (Rev. 2-89)  
Prescribed by ANSI Std. Z39-18  
298-102

## FOREWORD

Opinions, interpretations, conclusions and recommendations are those of the author and are not necessarily endorsed by the U.S. Army.

N/A        Where copyrighted material is quoted, permission has been obtained to use such material.

N/A        Where material from documents designated for limited distribution is quoted, permission has been obtained to use the material.

☒ Citations of commercial organizations and trade names in this report do not constitute an official Department of Army endorsement or approval of the products or services of these organizations.

X In conducting research using animals, the investigator(s) adhered to the "Guide for the Care and Use of Laboratory Animals," prepared by the Committee on Care and use of Laboratory Animals of the Institute of Laboratory Resources, national Research Council (NIH Publication No. 86-23, Revised 1985).

X For the protection of human subjects, the investigator(s) adhered to policies of applicable Federal Law 45 CFR 46.

X In conducting research utilizing recombinant DNA technology, the investigator(s) adhered to current guidelines promulgated by the National Institutes of Health.

X In the conduct of research utilizing recombinant DNA, the investigator(s) adhered to the NIH Guidelines for Research Involving Recombinant DNA Molecules.

X In the conduct of research involving hazardous organisms, the investigator(s) adhered to the CDC-NIH Guide for Biosafety in Microbiological and Biomedical Laboratories.

Peter V. Hanschke 1/20/00  
PI - Signature Date

Annual Report**Bone Factors Regulating the Osteotropism of Metastatic Breast Cancer**

Principal Investigator:

**Peter V. Hauschka, Ph.D.**Senior Research Associate, Department of Orthopaedic Surgery  
Children's Hospital, Boston, MA 02115Table of Contents

<u>Section</u>	<u>Page</u>
Front Cover Page (Form)	
Report Documentation Page (Form SF298)	
Foreword (Form)	
Table of Contents	4
Abstract (Repeated from Face Page)	5
Introduction	5
Body of Report	6
Overview of "Statement of Work"	6
Progress on Technical Objective #1	6
Methods	6
Results	7
Progress on Technical Objective #2	11
Methods	11
Results	11
Progress on Technical Objective #3	18
Methods	18
Results	18
Progress on Original Milestones	21
Key Research Accomplishments	21
Reportable Outcomes	22
Conclusions	22
References	25
Appendix	28
Final Report Data	32

**Abstract (Repeated from Face Page)**

The specificity with which breast cancer metastasizes to the human skeleton is currently unexplained. In this project we have attempted to identify the critical molecular interactions and cellular processes responsible for this osteotropism (homing and metastatic growth in bone) of breast cancer. The central hypothesis is that malignant breast adenocarcinoma cells subvert the cooperative paracrine interactions between normal bone cells, endothelial cells, and the extracellular matrix in order to establish metastatic foci. Our data combined with published reports show that invasive breast adenocarcinoma cells mimic certain hallmarks of the osteoblast phenotype, possibly explaining the facile growth and survival of these tumor cells in bone. Analysis of model human tumor lines, and derivative sublines recovered from bone metastases in nude mice, reveals a correlation between the degree of malignancy and the inappropriate expression of the "bone specific" transcription factor CBFA1. The osteoblast mimicry by the tumor cells may be explained by CBFA1 expression; normal breast tissue does not express CBFA1. Osteoblast factors also have a paracrine effect on altering the phenotype of breast adenocarcinoma cells in the bone microenvironment.

**Introduction**

Osteotropism of metastatic breast cancer (i.e., the predisposition of malignant adenocarcinoma cells to lodge in bone and establish painful osteolytic metastases) is the focus of this Project, and has direct relevance to three of the goals of the USAMRMC/DOD Breast Cancer Research Program:

1. We are defining changes in cell and molecular functions accounting for the development and progression of breast cancer.
2. The findings could be of potential use in guiding diagnosis, prevention, and treatment.
3. The research targets a major economic element (the costs of hospitalization and clinical management of skeletal morbidity) and could lead to more cost-effective health care delivery.

Malignant breast cancer cells exploit the normal bone homeostatic mechanisms in 3 ways for their own benefit: 1) secretion of PTH-rp and other factors stimulates osteolysis, creating space for metastatic tumor growth; 2) growth factor release during pathologic osteolysis drives adenocarcinoma cell proliferation; and 3) angiogenesis caused by osteolysis provides increased blood flow and a plentiful supply of nutrients to growing metastases. Partial mimicry of the osteoblast phenotype by malignant breast adenocarcinoma cells<sup>1-3</sup> is hypothesized to allow the expropriation of normal osteoblast-osteoclast signaling pathways for the exclusive benefit of growing tumor metastases in bone. Release of growth factors and rapid angiogenesis occurs in the immediate vicinity of an active osteoclast.<sup>4,5</sup> Osteoblast-derived bone sialoprotein (BSP), osteopontin (OPN), and osteocalcin (OC) are required for osteoclast attachment and activation,<sup>5-10</sup> and expression of BSP by primary human breast adenocarcinoma is a recent predictor of metastasis.<sup>3</sup> Little is known about the regulation of such events. We are using *in vitro* and *in vivo* models to characterize this important pathological mimicry by malignant breast cancer cells.

Our ongoing and planned experiments should provide insight into the biology of painful bone metastasis in invasive breast cancer, offering hope for innovative therapy. Given the unchanging survival outlook for patients with metastatic bone disease, any significant improvement in the quality of remaining life would stand as an important therapeutic effect.

## Body of Report

### Overview of "Statement of Work"

There are three Technical Objectives comprising the original Statement of Work for this project. On the following pages, results for the entire project period have been organized according to these Technical Objectives and the specific Tasks within each section on which progress has been made. Where appropriate, difficulties preventing the completion of individual Tasks are addressed.

#### Progress on Technical Objective #1

**Technical Objective 1:** Examine the regulation of malignant human breast adenocarcinoma cell lines by bone extracellular matrix proteins and by normal osteoblasts *in vitro*.

**Task 2:** Develop baseline data on cell proliferation and gene expression by the MCF-7 human breast adenocarcinoma cell line cultured on various bone ECM constructs. Follow this with a parallel study of the more malignant estrogen-independent MDA-231 line.

**Task 3:** Using Expt.#2, where normal osteoblasts are included in co-cultures with breast adenocarcinoma cells, study the process of osteoblastic mimicry. That is, define the expression of bone matrix proteins (BSP, OPN, OC) by adenocarcinoma cells. With trans-well chambers, test the role of cell-matrix and cell-cell (adenocarcinoma-osteoblast) contact in the regulation of gene expression and proliferation.

**Methods:** The primary analysis involves comparison of adenocarcinoma cell gene expression and proliferation in the presence and absence of normal osteoblasts. A proven method for this interactive analysis of soluble factors is co-culture, either in the trans-well format, where tumor cells growing on a semi-permeable membrane are placed in proximity to the osteoblast monolayer culture, or it direct cell-cell contact, allowing the analysis of cell-matrix and cell-cell (adenocarcinoma-osteoblast) contact in the regulation of gene expression and proliferation. Our prime focus is to define the regulation of adenocarcinoma gene expression by bone factors. A secondary focus is modulation of the osteoblast phenotype by the tumor cells.

**Cells:** Human Breast Cancer (BrCa) Cell Lines: **MCF-7** (estrogen-dependent human breast adenocarcinoma); **MDA-MB-231** (estrogen-independent human breast adenocarcinoma, more aggressively malignant and metastatic than MCF-7); **M1**, **M2**, and **Blast-4** sublines of MDA-MB-231 recovered, cultured, and passaged during this project from individual bone metastases in nude mice; C5 (another MDA-MB-231 subline recovered by M. Tondravi); **HMEC** (normal human breast epithelial cells); **hFOB** (human fetal osteoblasts); and **MC3T3** (clonal mouse osteoblasts).

**RNA/RT-PCR** RNA extraction was performed using RNeasy Mini Kit (Qiagen, CA). Primers were synthesized to unique regions in the osteocalcin, CBFA1, and GAPDH human cDNA sequences reported in the Genbank (Table 1). PCR reaction cycle conditions consisted of denaturation at 93°C for 60s, primer annealing at 61°C for 60s, extension at 72°C for 60s, and a final extension at 72°C for 7 min in a MJ Research (PTC-100) programmable thermal-cycler. Samples were amplified for 25 cycles. UV photography and densitometry of ethidium bromide stained gels



was performed. Semi-quantification was achieved by comparing the amplicon intensities of both CBFA1 and osteocalcin to GAPDH.

**Quantitative RT-PCR:** This method has only recently become available in our institution. We have now designed and synthesized TaqMan primer sets for human osteocalcin as an initial test of this technology. Using a Perkin-Elmer instrument, we have measured a  $C_T$  value of 13.3 for an input of only 40 femtograms of human osteocalcin cDNA target. This means that a single molecule of osteocalcin cDNA (about  $2 \times 10^{-17}$  g cDNA) in an RT sample prepared from cellular RNA should have a  $C_T$  of about 28 cycles, well within the limit of 35-40 cycles for a typical run. As warranted we have planned to expand this approach to CBFA1 and other bone genes to complete the analysis of the adenocarcinoma expression profile.

**Gel Shift Assay (EMSA):** Nuclear extracts are prepared by 0.45M KCl extraction. Electrophoretic mobility assays are performed using standard conditions.<sup>11</sup> DNA binding reactions will be performed as described.<sup>12,13</sup> Nuclear extracts are incubated with 10 fmol of the  $^{32}$ P labeled CBFA1 binding site consensus oligonucleotide (5'-CGAGTATTGTGGTTAATACG-3') as the probe and nonspecific competitor DNA. Protein-DNA complexes are resolved on a 4% non-denaturing polyacrylamide gel. Anti-sera supershift experiments will utilize polyclonal anti serum directed to a CBFA1 peptide containing 17 amino acids<sup>13</sup> or preimmune serum (control). (Gift of Dr. Scott Hiebert, Vanderbilt Medical Center Nashville TN/ and Oncogene Research Products, Cambridge, MA).

**Osteocalcin Radioimmunoassay (RIA):** Human osteocalcin concentrations in conditioned medium samples (1,3,5,7 days) were measured by radioimmunoassay using rabbit anti-monkey osteocalcin (first antibody) and goat anti-rabbit IgG (second antibody).

**Osteocalcin Western Blot:** This is a totally new method which we developed during the course of this project. It enabled us to solve the difficult question of whether osteocalcin expression, at the protein level, is detectable in breast cancer cells, and whether it changes in response to osteoblast factors. The technical difficulty is that osteoblasts are heavy producers of osteocalcin, so it would be impossible to perform this experiment with human BrCa cells and human osteoblasts. However, we developed two antibodies that recognize osteocalcins of the human (primate) and mouse sequence in a species specific fashion. Therefore, as shown in the western blots below, a coculture of mouse MC3T3-E1 osteoblasts with human BrCa lines can be examined for bidirectional regulation of osteocalcin expression by both osteoblasts and tumor cells.

**General Cell Extraction:** Cultures were rinsed with PBS, scraped into buffer and collected by centrifugation. Cells were lysed with hypotonic buffer in the presence of protease inhibitors. Nuclei were collected by centrifugation and resuspended in hypertonic buffer and protease inhibitors. Nuclei were extracted on ice, insoluble material was cleared by centrifugation and soluble proteins were stored at  $-80^\circ\text{C}$ . Nuclear extracts (30  $\mu\text{g}$  protein per lane) were resolved with 8% SDS-PAGE and electroblotted onto nitrocellulose membrane. Membranes were incubated with a 1:100 dilution of antibody in Tris-buffered saline containing 1% BSA. Affinity purified antibodies specific for CBFA1 were used in these studies. Membranes were incubated with secondary antibody, and protein bands were visualized by streptavidin-HRP chemiluminescence (Pierce, IL).

**Results:** Human breast adenocarcinoma cell lines have been established in the laboratory and examined by RT-PCR, Northern analysis, immunohistochemistry, and radioimmunoassay to establish

the level of expression of osteoblast-specific genes. The focus has been maintained on the MDA-MB-231 adenocarcinoma line, based on our success in obtaining osteolytic metastases with this same line in nude mice (see Technical Objective #3 below).

**RT-PCR for CBFA1.** (Figure 1) Semi-quantitative PCR was performed comparing the expression of CBFA1 and osteocalcin between HMEC, MDA-MB 231 cell line, and M1. These results were normalized to GAPDH for each sample. There was a two fold increase in both CBFA1 (375bp) and osteocalcin (294bp) in the M1 clone (lanes 5 and 2) compared to the original MDA-MB 231 cell line (lanes 4 and 1). There was no evidence of CBFA1 in the HMEC (lanes 3 and 6) cell line. PCR product identity was confirmed by producing products of expected size.. The CBFA1 amplicon was excised from the gel, purified and underwent automated sequencing to further confirm its identity.

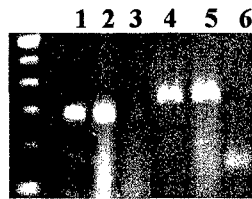


Figure 1

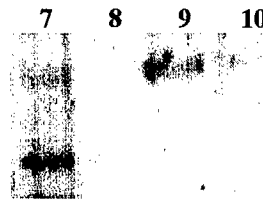


Figure 2

**Western Blot for CBFA1:** (Figure 2) We examined nuclear extracts from the MDA-MB 231 cell line, HMEC line, and M1 subclone, and MG-63 osteosarcoma cell line (positive control) using antibody to CBFA1. CBFA1 was detected in all of these cell lines except the normal breast line HMEC (Lane 8). A 60-65 kDa species was present in the MDA-MB 231 cells (Lane 10) and M1 subclone (Lane 9). The species was present at a two-fold greater band density in the M1 clone (Lane 9). The MG-63 human osteosarcoma cells (Lane 7) expressed the 60-65 kDa form as well as a prominent 46 kDa form that was not seen in the breast cancer cell lines.

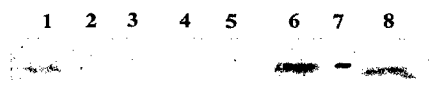
**Supershift and Electromobility Shift Assays (EMSA) for CBFA1:** The possibility of CBFA1 expression was analyzed by EMSA on nuclear extracts prepared from cultured breast adenocarcinoma and normal cell lines. CBFA1 transcription factor activity is detected by retarding the mobility of a specific radioactive target oligonucleotide containing the CBFA1 binding site that is common to a number of osteoblast-specific gene promoters. The labeled oligonucleotide was shifted to slower mobility by putative CBFA1 activity in MDA-MB-231, M1, M2, and Blast-4 cell extracts. HMEC extracts failed to show CBFA1 activity. MG-63 osteosarcoma served as a positive control for CBFA1, and the EMSA assays showed the expected competition with excess cold target oligonucleotide. Importantly, the specific antibody to CBFA1 caused further retardation of the label (supershifting), allowing us to conclude that active CBFA1 had been upregulated in the BrCa cells. This finding provides a likely explanation for the osteoblastic mimicry by BrCa cells.

**Breast Cancer Cells Express Increased Levels of Osteocalcin Compared to Normal Breast Tissue:** The rationale for examining osteocalcin expression is that this is one of the "osteoblast specific genes" that is transcriptionally regulated by CBFA1. Because CBFA1 is upregulated in the tumor lines, we have analyzed the conditioned media from tumor and normal breast cells for osteocalcin by two methods. Human osteocalcin concentrations in conditioned medium samples of the MDA-MB 231 cells line and HMEC cell lines were measured by radioimmunoassay using a specific osteocalcin antibody. Data are reported as ng osteocalcin/ml of medium. Medium (with

10% FCS) served as the baseline control, and this accounts for the background which is 13 ng/ml of cross reacting bovine osteocalcin contributed by the 10% serum (FCS). MDA-MB-231 cells expressed increasing amounts of osteocalcin with increasing confluence. The HMEC cell line did not produce osteocalcin when compared to medium controls. However, other less osteotropic cell lines have not yet been examined to determine if there is an association between osteocalcin production and ability to metastasize to bone. Future experiments will examine less osteotropic breast cancer cell lines, non-osteotropic and non breast malignancies (i.e. colon cancer), as well as the M1 and the LC115 bone metastasis derived cell lines, in order to corroborate the apparent positive correlation between high osteotropism and expression of CBFA1 and osteocalcin.

#### Western Blot Analysis of Osteocalcin Expression:

**Figure 3.** Species specificity of osteocalcin detection in conditioned media. The upper blot detects the 6kDa mouse osteocalcin protein in lanes 1 (mouse MC3T3 osteoblasts), 6 (MC3T3 cultured in the mineralizing promoter beta glycerolphosphate), and 8 (mouse osteocalcin standard 5ng). No human osteocalcin in the human BrCa cell line MCF-7 is detected in lanes 3 or 4, nor is the primate osteocalcin standard in lane 7 detected (5 ng monkey osteocalcin). In the lower blot, the human osteocalcin is prominent in the MCF-7 conditioned medium when cocultured with mouse osteoblasts (lane 7), but only weakly detectable in the tumor cells alone (lane 5, 6). Mouse osteoblasts (lane 8) show no background expression of human osteocalcin cross-reactivity.



#### **Western Blot (Goat anti-mouse OC antibody)**

1: 3T3 2/19; 2: 3T3 2/19 + MCF 2/24  
3: MCF 2/24; 4: MCF 2/24 + GP;  
5: 3T3 2/19+MCF2/24+GP; 6: 3T3 2/19+GP  
7: Marker; 8: 5ng OC + alpha-MEM

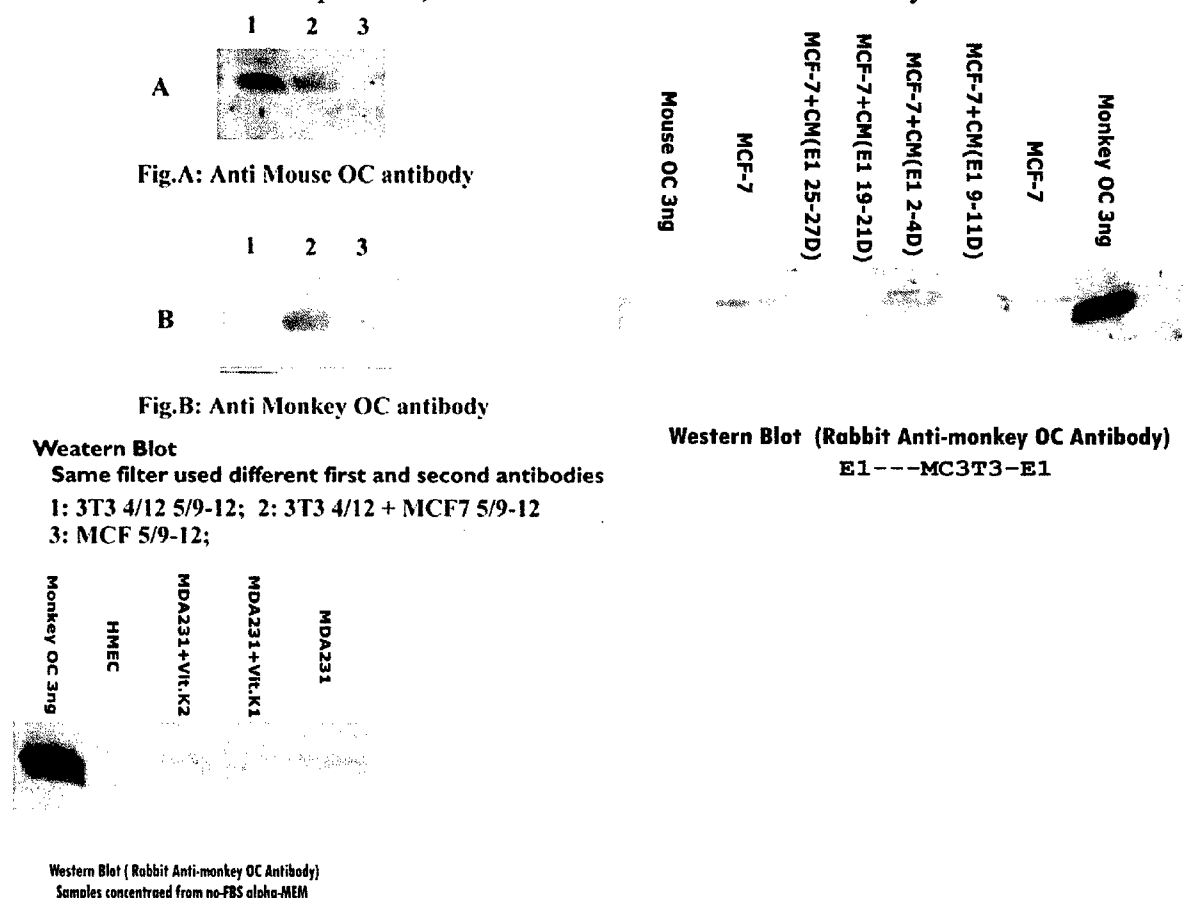


#### **Western Blot**

##### **Rabbit anti-monkey OC antibody**

1: 3 ng monkey OC      2: Marker  
3: 3T3 2/19+GP; 4: 3T3 2/19 + MCF 2/24 +GP  
5: MCF 2/24+GP; 6: MCF 2/24;  
7: 3T3 2/19+MCF2/24; 8: 3T3 2/19

**Figure 4.** Other western blots showing the reproducible detection of human osteocalcin in the conditioned media of MCF-7 cells, particularly when cocultured with osteoblasts (MC3T3) or with CM (concentrated conditioned media) from osteoblasts. The normal breast epithelium (HMEC) shows no osteocalcin expression, while MDA-MB-231 shows extremely low levels.



The western blot data for osteocalcin are in need of corroboration by RIA and quantitative RT-PCR (TaqMan) analysis of gene expression. It should be noted that while the CBFA1 expression levels appear to correlate with the malignancy of the BrCa cell type (M1 > MDA-MB-231 > MCF-7 > HMEC), there is not a similar correlation with osteocalcin protein levels, where MCF-7 > MDA-MB-231 > A2, HMEC. Some of the explanation may lie in the proteinases secreted by each of the cells, as osteocalcin is very susceptible to proteolytic fragmentation which destroys its immunoreactivity.

**Task 4:** Study the action of hOP-1 in regulating adenocarcinoma cells.

Tumor cell culture with hOP-1 showed decreasing osteocalcin after day 7 by RIA analysis. Prior to publication, this important observation needs to be repeated, and we will also examine proliferation effects, as hOP-1 also appears to slow the tumor cell growth in vitro. In designing the future experiments, we have obtained two new sources of human osteoblasts [FOB (Mayo Clinic)

and NHOst (Clonetics)], along with the normal human breast epithelial line, HMEC (which is negative for osteocalcin).

### **Progress on Technical Objective #2**

**Technical Objective 2:** Study the bidirectional interaction between breast adenocarcinoma cells and bone cells (osteoblasts and osteoclasts) *in vitro*.

**Task 5:** Months 6-12. Establish the 3-D co-culture model (Expt. #3) initially with MCF-7 adenocarcinoma cells and human MG63 osteoblast-like osteosarcoma cells, testing the variables of cell density and time of contact between cell clusters on proliferation and gene expression of the two cell types. Adapt normal mouse MC3T3-E1 osteoblasts to growth in the 3-D gelatin gel configuration in order to examine the actions of the adenocarcinoma cells in perturbing the normal osteoblast phenotype. Follow this with dose-dependence studies of 17- $\beta$  estradiol and hOP-1 effects in the model.

**Task 6:** Months 9-15. Establish the resorbing calvaria co-culture model (Expt. #4), initially comparing standard resorption stimuli (PTH, PGE<sub>2</sub>) with varying numbers of MCF-7 adenocarcinoma cells and MCF-7 conditioned medium. Use blocking antibodies, peptide antagonists of PTH-rp, and indomethacin to define the nature of the resorptive stimulus originating from the adenocarcinoma cells.

**Task 7:** Months 12-18. Investigate the effects of resorbing bone culture medium on MCF-7 and MDA-231 adenocarcinoma cells, using appropriate controls for the carry-over of PTH or PGE<sub>2</sub>. Focus on differentially regulated cell proliferation and the expression of genes which are known to be correlated with metastasis. Study the comparative effects of 17- $\beta$  estradiol, tamoxifen, and hOP-1 on the MCF-7 and MDA-231 responses.

**Methods:** Add tumor cells to established cultures of normal osteoblasts for direct contact co-culture, and search for changes in gene expression and proliferation of the tumor cells which require cell contact or labile products of living osteoblasts.

**Results:** The standard method adopted for co-culture studies was to plate MC3T3 mouse osteoblasts, allow them to attach, and then to seed the surface with tumor cells (either MCF-7 or MDA-MB-231) with a delay of between 0 and 45 days. When plated with minimal delay, we observed that both tumor cell types would rapidly attach and outgrow the osteoblasts. In the case of MCF-7, the osteoblasts were adversely affected and a moderate fraction began to undergo detachment and apoptosis within several days of contact. However, MDA-MB-231 were observed to require as much as 5-7 days before causing osteoblast apoptosis, and in this case there was evidence of severe cytotoxicity with virtually 100% of the osteoblasts being killed.

Very different results were obtained for tumor cells plated on established, mineralized osteoblast cell layers. In this situation, both tumor cell types required many days to attach and proliferate to the point where small clonal colonies of 5-10 cells could be distinguished. Because the morphological features of MCF-7 made these colonies very easy to identify, most of the long-term experiments involved MCF-7 rather than MDA-MB-231. Importantly, there was clear evidence of paracrine effects of the tumor cells on the osteoblasts. Those osteoblasts that were in immediate proximity or contact with an MCF-7 colony were found to survive and appeared to produce increased amounts of mineralized matrix. However, a high percentage of osteoblasts that were distant from the tumor cells underwent apoptosis.

Trans-well co-culture experiments with of MDA-MB-231 cells and MC3T3-E1 cells were difficult and therefore abandoned. The principal problem is the rapid growth of the tumor cells and the resulting nutrient depletion and medium acidification which adversely affects the slower-growing osteoblasts. These experiments were discontinued in favor of direct co-culture. It required much trial and error to determine the appropriate plating densities and culture conditions. The optimal conditions for co-culture are established for 24-well plates: 1) plate  $1.2 \times 10^5$  MC3T3 cells (day 0); 2) overlay  $1.2 \times 10^4$  MCF-7 cells when osteoblasts cover 80% of the culture surface (day 2); 3) fix at day 4 or later for immunohistochemistry.

Table 1. Immunohistochemistry of Breast Adenocarcinoma (MCF-7) and Osteoblastic (MC3T3) Cells

Antigenic Marker	Cells Cultured Alone		Cells Co-Cultured (2d)	
	MCF-7	MC3T3	MCF-7	MC3T3
human cytokeratin-18	+++	—	++++	—
bone sialoprotein (BSP)	++	—	+++	—
osteopontin (OPN)	++	—	+++	—
osteocalcin (OC)	+	±	++	±
alkaline phosphatase	—	+++	±	+++
non-immune serum control	—	—	—	—

**Immunohistochemistry:** Table 1 shows the results of antibody staining of breast adenocarcinoma (MCF-7) and osteoblastic (MC3T3) cells alone and in co-culture. The human-specific cytokeratin-18 antibody clearly distinguishes the epithelial adenocarcinoma cells from the murine osteoblasts, as expected. The osteoblasts are selectively stained by alkaline phosphatase (BCIP reaction), and weakly for osteocalcin, as expected for this early time point in the culture (day 4). What is important about the data in Table 1 is the relatively strong expression of the bone cell markers by the MCF-7 cells, and the observation that the staining increases when the MCF-7 tumor cells are co-cultured with the MC3T3 osteoblasts. The growth behavior of the MCF-7 cells in co-culture is in isolated clonal clusters containing 5-50 cells which are surrounded by flattened osteoblasts. The density of the positive immunohistochemical staining for the proteins such as BSP and OPN is greater at the periphery of these clusters, suggesting either a contact-dependent regulation of gene expression, or a possible uptake of osteoblast-produced proteins by the tumor cells. These possibilities will be discriminated by future experiments employing in situ hybridization and species-specific immunostaining.

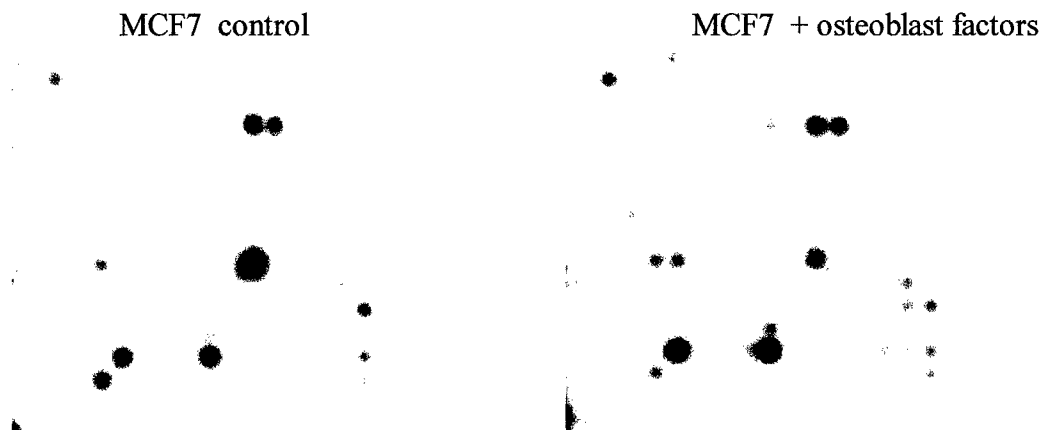
**Bidirectional Interactions of Tumor Cells and Osteoblasts:** It is noteworthy that when larger numbers of MCF-7 cells are plated on the osteoblasts (i.e., when the ratio of tumor cells/osteoblasts is greater than 1/2, then the osteoblasts lift off the culture surface and die within 3 days. Even if the medium is buffered by HEPES to prevent overt acidification by the metabolically active tumor cells, rounding and detachment of the osteoblasts is only delayed by 1-2 days.

Conditioned medium experiments showed that both MCF-7 and MC3T3 cells produce attachment factors which increase each other's attachment to plastic in serum-free conditioned alpha-MEM. However neither of the cell types can survive beyond 48-72 hr in these conditioned media,

presumably because of the absence of serum growth factors. At this time, we have no evidence of overt cytotoxic factors being produced by either cell type for action on the other.

Gene Expression Array Analysis of BrCa Regulation by Osteoblasts: The major new development of commercially produced Gene Expression Arrays in the past 6 months has provided a comprehensive technique for profiling the total phenotype of normal and cancer cells. These new arrays allow the simultaneous analysis of thousands of genes at a time, rather than the few genes to which most previous studies had been restricted. We have utilized the new advanced arrays of 1176 known cDNAs produced by Clontech. Initial work has proven the power of this approach in analyzing the effects of osteoblast conditioned medium on gene expression by MCF7 cells after 24 hr treatment. The small region (about 200 genes) of the array shown below (Fig. 5) demonstrates the density of valuable information that can be obtained. Each spot contains 10 ng of cDNA from a known human gene, and the black density in the autoradiogram is proportional to the expression level of mRNA for each gene in the MCF7 cell culture. Note that the expression of many genes is not affected by the added osteoblast factors, whereas others are increased or decreased.

**Figure 5.** Clontech Human Gene 1.2 Array - sector A. Left side is hybridized with  $^{32}\text{P}$ -labeled cDNA from control MCF7 cells; right side is MCF7 + osteoblast CM; red arrows highlight regulated genes (+, increased by osteoblast factors; -, decreased)



We are now in the process of developing a list of the important genes that are turned up or down by osteoblast-derived factors. This provides a window into the vast and interconnected responses of BrCa cells to the bone microenvironment. A complementary set of experiments is in progress to analyze the effects of the BrCa cell-derived factors on osteoblasts and osteoclast precursors.

We have performed many experiments to date, and some of the more clearcut changes in gene expression are shown in the table and list below:

Human Breast Adenocarcinoma MCF-7 Cell Line:

48 hr Gene Regulation by Osteoblast Factors

Up-regulated on Human 1.2 Array:

A03h: G1/S-specific cyclin D1 (DDND1), cyclin PRAD1, bcl-1 oncogene  
 A13j: cyclin-dependent kinase regulatory subunit 1(CKS1)  
 A13I: cdc2-related protein kinase PISSLRE  
 A03I: prothymosin alpha (ProT-alpha, PTMA)  
 B08e: transferrin receptor (TFRC); CD antigen  
 B10n: guanine nucleotide-binding protein G-i/G-s/G-t beta subunit 2; transducin beta 2 subunit2  
 C12a: serine/threonine protein phosphatase PP1-alpha isoform (PP-1A) (also see in Human 1.2II)  
 C06I: apoptosis regulator bax  
 C14I: NIP3 (NIP3)  
 C04j: rac-alpha serine/threonine kinase (rac-PK-alpha); protein kinase (PKB); c-akt; akt1  
 C09j: IEX-1L anti-death protein; PRG-1; DIF-2  
 C05k: cytoplasmic dynein light chain 1 (HDLC1); PIN  
 C05I: DNA topoisomerase II alpha (TOP2A)  
 D10k: ADA3-like protein  
 D11m: helix-loop-helix protein; DNA-binding protein inhibitor ID-2  
 E04b: nuclease-sensitive element DNA-binding protein (NSEP)  
 E08c: trans-acting -cell specific transcription factor GATA3  
 E02e: cAMP-dependent transcription factor ATF-4; CREB2  
 E08e: guanine nucleotide-binding protein G-s alpha subunit (GNAS);  
 E11n: microsomal glutathione S-transferase 12 (GST12; MGST1)  
 F07a: natural killer cell enhancing factor (NKEFB)+ TSA; TDPX1  
 F07d: platelet-derived growth factor B subunit precursor (PDGFB; PDGF2)  
 F13m: cathepsin D precursor (CTSD)  
 F05n: metalloproteinase inhibitor 1 precursor (TIMP1); EPA;

Up-regulated on Human 1.2-II:

CD 81 antigen; 26-Kda cell surface protein TAPA-1  
 Dek protein  
 LANP cerebellar leucine rich acidic nuclear protein  
 RNA -binding protein fus/tsl  
 Ras-related protein RAB-7  
 Clathrin heavy subunit 1(CLH-17);KIAA0034  
 Brain-form phosphoglycerate mutase;PGAM-B; BGP-dependent PGAM  
 6-phosphofructokinase, type c (phosphofructokinase 1) (phosphohexokinase)  
 Pyruvate kinase M2 isozyme (PKM2)  
 Ribosomal protein S6 kinase (EC 2.7.1-) (S6K) (P70-S6K)  
 PS2 protein precursor; HP1.A; breast cancer estrogen-inducible protein (BCEI); TFF1;PNR2  
 Contains proprietary or unpublished data  
 Limit distribution appropriately



Natriuretic peptide precursor B

Serine/threonine protein phosphatase PP1-alpha 1 catalytic subunit (PP-1A)

Guanine nucleotide-binding protein beta subunit-like protein 12; receptor of activated protein kinaseC1 (RACK1)

calgizzarin; S100 C protein;MLN70

astrocyte glial fibrillary acidic protein (GFAP)

Down-regulated on Human 1.2 Array:

A04c: transforming growth factor -beta signalling protein 1(BSP1)

A02h: G2/mitotic-specific cyclin B1 (CCNB1)

A14j: cyclin-dependent kinase regulatory subunit (CKS2)

B06g: NCK melanoma cytoplasmic src homolog (HSNCK)

C04e: hint protein; protein kinase C inhibitor (PKCII)

C01h: CD40 receptor-associated factor 1 (CRAF1)

C12I: bcl-2 interacting killer (BIK); BP4; BIP1

D14j: activated RNA polymerase II transcriptional coactivator p15; PC4

D02n: 60s ribosomal protein L6 (RPL6)

E04c: nuclear factor NF45

E03d: transcription initiation factor TFIID 31kD subunit; TAFII31; TAF2G

E05k: monocyte chemoattractant protein 1 receptor (MCP-1RA); CCCR2; CCR2

F04b: heat shock 90kD protein A; HSP86

F14e: amphiregulin (AR); CRDGF

F13f: thymosin beta-10 (TMSB10; THYB10); PTMB10

F13j: thymosin beta 4; FX

F13k: proteasome activator HPA28 subunit beta

F02l: proteasome component C3; macropain subunit C3

Down-regulated on Human 1.2-II:

LGALS3, MAC2 (Galectin-3, MAC-2 antigen, IGE-binding protein, 35KD lectin)

Caltractin isoform 1; centrin

Ferritin heavy chain (FTH1);FTHL6

60s ribosomal protein L22 (RPL22); Epstein-Barr virus small RNA associated protein (EBERassociated protein; EAP); heparin binding protein 15 (HBP15)

F07h: calcium-dependent protease small (regulatory) subunit, calpain, calcium-activated neutral proteinase (CANP)

F03d: calcium-binding protein ERC-55 precursor

F08c: lipoprotein-associated coagulation inhibitor

F07I: huntington interacting protein (HIP2)=ubiquitin

F02j: calmodulin

F04k: G-alpha interacting protein (GAIP)

F12j: myristoylated alanine-rich C-kinase substrate (MARCKS)

Human Breast Adenocarcinoma MDA-MB-231 Cell Line:

48 hr Gene Regulation by Osteoblast Factors

Contains proprietary or unpublished data

15

Limit distribution appropriately

Up-regulated on Human 1.2 Array:

A07l: 40s ribosomal protein S19 (RPS19)  
 A06m: erythrocyte glucose transporter 1 (GLUT1)  
 C09j: IEX-1l anti-death protein; PRG-1; DIF-2  
 C13l: replication factor C 38-kD subunit (RFC38); activator 1 38-kDa subunit  
 D08j: hypoxia-inducible factor 1 alpha (HIF1 alpha); ARNT-interacting protein; MOP1  
 F11m: MMP16; MT-MMP3; MMP-X2

Down-regulated on Human 1.2 Array:

A03b: EB1 protein  
 A03l: prothymosin alpha (ProT-alpha; PTMA)  
 B06k: serine kinase  
 B13m: ras-related C3 botulinum toxin substrate 1; p21-rac-1;  
 C02l: calcium-dependent protease small (regulatory ) subunit; clpain; calcium-activated neutral proteinase (CANP) (also see in MCF-7 Human 1.2II)  
 C05k: cytoplasmic dynein light chain 1 (HDLC1); PIN  
 C05l: DNA topoisomerase II alpha (TOP2A)  
 E04b: nuclease-sensitive element DNA-binding protein (NSEP)  
 E02e: cAMP-dependent transcription factor ATF-4; CREB2  
 E05e: putative transcription activator DB1  
 E13g: alpha-1 catenin (CTNNA1);  
 E08h: CD44H; PGP-1; HUTCH 1  
 F10g: MIP2-alpha; GRO-beta  
 F13m: cathepsin D precursor (CTSD)  
 F05n: metalloproteinase inhibitor 1 precursor (TIMP1); EPA;  
 F03n: endothelial plasminogen activator inhibitor-1 precursor (PAL1; PLANH1)

Table 2 indicates a partial list of the important genes for which the expression level is turned up or down at least 2-fold by osteoblast-derived factors in the MCF7 cell line, a well studied breast adenocarcinoma model.

This type of analysis provides a clear window into the vast and interconnected responses of BrCa cells to the bone microenvironment. A complementary set of experiments is in progress to analyze the effects of the BrCa cell-derived factors on osteoblasts and osteoclast precursors.

**Table 2.** Regulation of MCF7 Adenocarcinoma Gene Expression by Osteoblast Factors

Increased Expression	Decreased Expression
<b><u>Growth Factor Related</u></b>	
PDGF-B	TGF-beta signaling protein (BSP1)
FGF-3/int-2	
VEGFR	
EGFR	
<b><u>Tumor Related</u></b>	
Estrogen receptor beta	Tumor suppressor protein DCC (colorectal cancer)
Estrogen related receptor gamma	
Prohibitin	
Bcl-1 oncogene (CCND1)	
Transforming protein rhoA (multidrug resistance)	
Cyclin dependent protein kinase 4 (CDK4)	
Transducer of erbB (TOB)	
P53 binding protein (TP53BP1)	
Manic fringe	
Elongation factor 1 alpha-1	
<b><u>Transcription Factors</u></b>	
GATA 3	Zinc finger GL1C
CREB2/ATF4	HNF-4-alpha
NSEB (YB-1)	C-1
	NFkappaB p52
<b><u>Signal Transduction</u></b>	
Protein kinase B (Akt1/PKB)	
MARCKS	
CAMP-dependent protein kinase (PRKAR1A)	
G protein Gi-beta	
Transducin beta-2	
Serine-threonine protein phosphatase PP-1A	
p21-rac-1	
<b><u>Cell Proliferation and Death/Apoptosis</u></b>	
NKEFB	Defender against cell death (DAD-1)
Ubiquitin protein ligase E2H10	Death domain containing protein (CRADD/RIADD)
M-phase inducer phosphatase (CDC25B)	
P55CDC	
<b><u>Extracellular Matrix</u></b>	
Metalloproteinase inhibitor TIMP-1	galectin-3 (laminin binding protein)
Integrin alpha-6	

The gene expression profiles for both MCF-7 and MDA-MB-231 show a number of important markers that have strong implications for their malignant phenotypes, including genes involved in growth factor stimulation, rapid proliferation and blocking apoptosis, transcriptional activation, efficient signal transduction, and tissue invasion. What is most significant in the data above is the patterns of gene expression that are then modulated by osteoblast-derived factors. The

action of osteoblast conditioned medium to turn up or down the expression of these tumor-related genes is unequivocal. What is not yet clear is whether a common set of genes is so regulated in every breast adenocarcinoma cell type, or whether each cell model has an intrinsically unique pattern. We are currently searching for these patterns.

### **Progress on Technical Objective #3**

**Technical Objective 3:** Establish human breast adenocarcinoma tumors at primary and skeletal metastatic sites by inoculation of athymic nude mice. Analyze the gene expression of tumor and bone cells in the recovered tissue. Test the activity of shOP-1 on primary tumor growth and on establishment and progression of skeletal metastases.

**Task 8:** Months 6-15. Using Expt. #5, inoculate MCF-7 cells and recover primary tumors from two subcutaneous sites (adjacent to fascia vs. calvaria). Measure tumor size and perform histology. Test the effect of shOP-1 on tumor growth and osteolysis in the calvarial site.

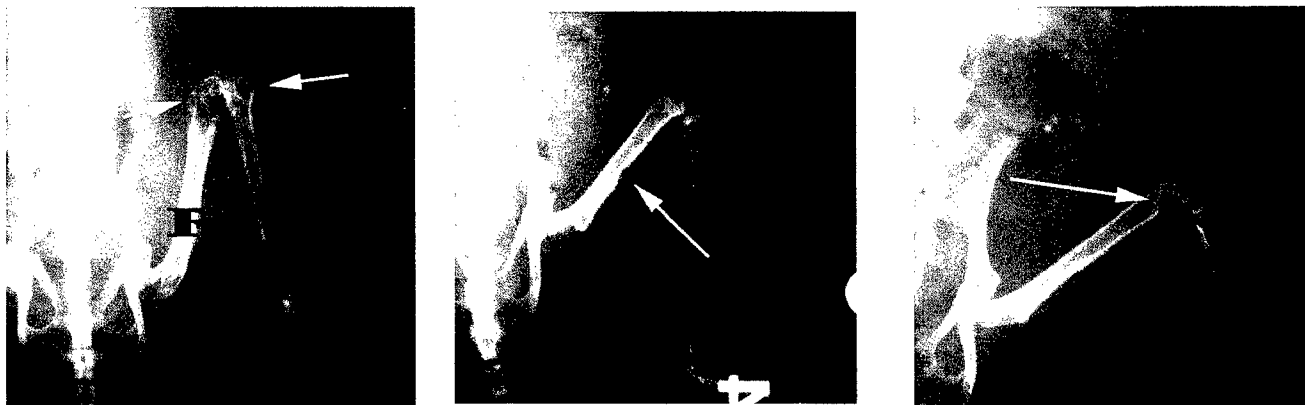
**Task 9:** Months 9-24. Establish the skeletal metastasis model (Expt. #6), first with MCF-7, and then with the more aggressive MDA-231 line. Perform a cell inoculation dose study and monitor osteolysis in developing skeletal metastases radiographically. Recover metastases for histology. Compare gene expression patterns by cells at the tumor/bone interface with tumor harvested from soft tissue (eg., lung).

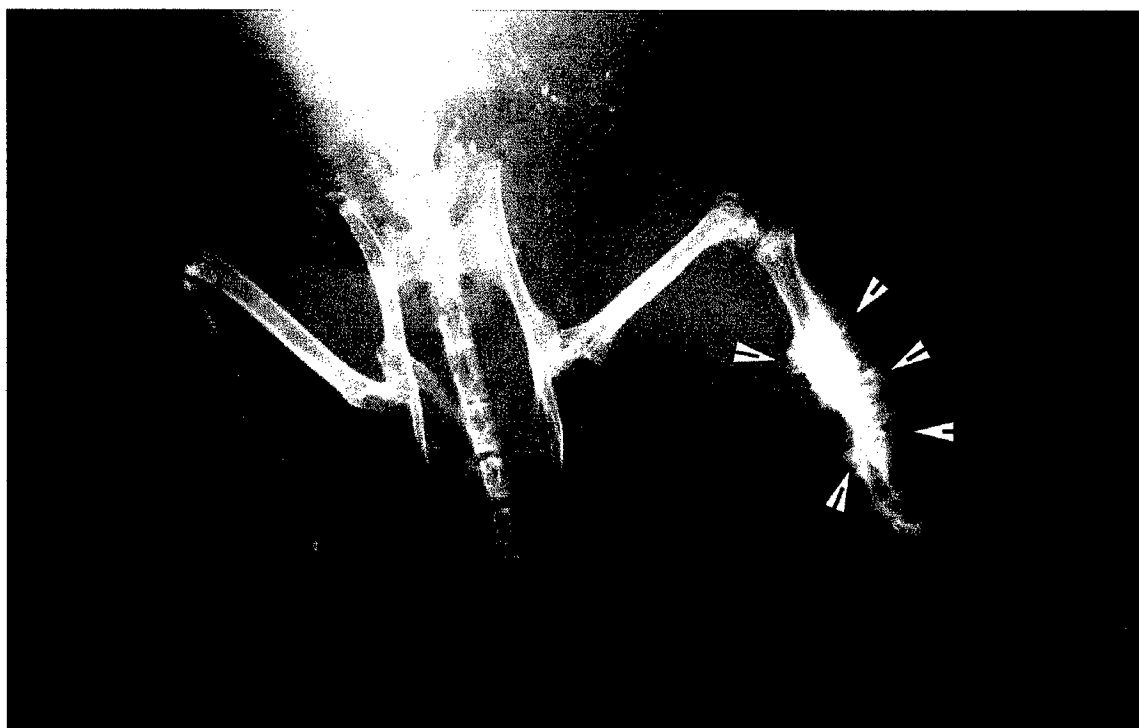
**Methods: Animals:** 7 week old, female Nu/Nu SCID mice (20-25g) were used following institutional guidelines for the use and care of laboratory animals. MDA-MB 231 cells were injected ( $1 \times 10^5$  cells/100 $\mu$ l) through the left second intercostal space into the left ventricle to produce bone metastases. Animals were radiographed bimonthly to detect the presence of lytic lesion. Bones with lesions were fixed in 4% paraformaldehyde and decalcified. Several lesions were isolated and subcultured to produce a breast cancer cell lines from a bone metastasis. These subclones were further characterized by immunostaining with cytokeratin 18 (mouse anti-human) to confirm both the human and glandular epithelial origin. Reinjection into mice was performed to confirm malignancy. **Immunohistochemistry:** Monoclonal or polyclonal antibodies to osteocalcin and CBFA1 and appropriate negative controls were employed. The sections were obtained from the distal femur and proximal tibia of mice. Archival human specimens from 10 patients with metastatic breast cancer to appendicular and axial sites were similarly examined. Fixed, paraffin embedded sections from each tumor were immunostained using an indirect avidin-biotin complex method. The appropriate concentrations of the primary antibodies against CBFA1 and osteocalcin were used (1:10 and 1:100, respectively).

### **Results:**

Over two years of experience with the mouse model for osteolytic metastasis of breast cancer has shown that only approximately 20% of mice develop metastases which are osteolytic by radiographic criteria. In about 2-5% of mice, multiple osteolytic sites are found. Three typical radiographs are shown in Fig. 6 below. The most common sites for osteolytic metastasis in the nude mouse were the femur and tibia. The radiolucent areas are evident, and these result from the osteoclastic osteolysis in the vicinity of the breast tumor metastasis.

**Figure 6.** Osteolytic metastases in mouse femur and tibia of 3 separate animals following injection of MDA-MB-231 human breast adenocarcinoma cells. After longitudinal monitoring of the lesions, animals were euthanized and the metastatic sites dissected, dispersed in sterile medium, and subcultured in vitro.





**Figure 7.** Strong osteoblastic reaction in response to an osteolytic metastasis of MDA-MB-231 cells. The white arrows show the dense, radioopaque new bone formed in the tumor site. The scalloped area (black arrows) results from tumor-mediated osteolysis. This lesion was the source of the Blast-4 cell line.

Recovery of Malignant Human Breast Adenocarcinoma Cells from Bone Metastases: Osteolytic bone metastases (and one osteosclerotic metastasis) were located radiographically in mice which had been injected by the intracardiac method with MDA-MB-231 cells. At sacrifice, 15 sites were dissected and curetted to obtain cells for tissue culture. Of these candidate cultures, 2 osteolytic ones (M1 and M2) grew vigorously and were verified to be sublines of the human MDA-MB-231 adenocarcinoma by human-specific cytokeratin-18 immunostaining which showed human, glandular epithelial origin. The osteosclerotic metastasis yielded a subline (Blast-4) which is also derived from the parental MDA-MB-231 line (Fig. 7).

Characterization of Osteoblast Gene Expression Patterns in Human Breast Adenocarcinoma Cells from Osteolytic and Osteosclerotic Metastases:

Immunohistochemical Investigation of Metastases Recovered from Mice: looked for CBFA1 in osteolytic tumor metastases and found positive evidence of the expression of this transcription factor in tumor foci, with little or no expression in mature bone itself. The CBFA1 expression by osteoblasts is primarily confined to the rapidly growing regions of the skeleton (growth plates, etc.). Osteocalcin expression was observed in the tumor foci more strongly than in the surrounding bone.

Gene Expression: The small size of the human breast adenocarcinoma metastases in mice makes it impossible to perform certain types of molecular analysis, so we have resorted to analyzing the adenocarcinoma cell lines (M1, M2, and Blast-4) recovered from individual metastases. M1 and

M2 are positive for CBFA1 expression RT-PCR (Fig.1), EMSA and Westerns (Fig.2). Northern analysis for CBFA1 has been problematic due to comigration of CBFA1 mRNA with the abundant 18s RNA. Blast-4 cells are positive for CBFA1 by EMSA and Western blotting. In summary, we have the parental line (MDA-MB-231), 3 recovered sublines (M1, M2, and Blast-4), and another MDA-MB-231 subline recovered by another laboratory (C5; M. Tondravi). All 5 of these cell lines express CBFA1, while normal breast cells (HMEC) do not.

Analysis of Archival Human Specimens: Archival human specimens have been obtained from 20 patients with metastatic breast cancer. The primary breast lesions and bone metastases have been obtained from the records of the Orthopaedic Oncology Service at the Massachusetts General Hospital. Paraffin block specimens from metastases in the femur have been obtained. Both osteocalcin and CBFA1 were noted to be positive for immunostaining in both archival human specimens and in experimentally induced animal models of bone metastasis. CBFA1 was seen primarily in the breast cancer cells within each metastasis, with minimal staining of the surrounding bone. Osteocalcin was seen in both the breast cancer metastasis and surrounding bone in both the archival human and animal specimens.

#### **Progress Summary for Milestones of the Original Research Plan:**

1. Demonstration of bone matrix-dependent and osteoblast-dependent regulation of malignant human breast adenocarcinoma cells. *(Accomplished and extended by gene expression arrays).*
2. Characterization of factors regulating osteoblastic mimicry by breast adenocarcinoma cells. *(Factors demonstrated, eg. CBFA1, but awaiting definitive proof that this is causative)*
3. Demonstration of adenocarcinoma-dependent modulation of the phenotype of normal osteoblasts. *(Work with osteoblasts still ongoing via gene expression arrays).*
4. Recovery of bone metastases of human breast adenocarcinoma from athymic nude mice in quantities suitable for histochemical and in situ hybridization studies of gene expression. *(The low frequency of metastases in the animal model did not provide sufficient tissue for statistically valid histochemical and in situ studies. Metastatic sublines were established from many lesions, providing a very promising collection of culturable materials for further study)*
5. Proof of anti-tumor efficacy of hOP-1 and shOP-1 in the experimental models. *(As above, the low frequency of metastases in the animal model did not provide sufficient tissue for statistically valid analysis of treatments. Accordingly, because of prohibitive projected costs and uncertain statistical power, this experiment was not performed).*

#### **Key Research Accomplishments:**

- Demonstration of gene expression of the bone-specific osteoblast lineage transcription factor CBFA1 in human breast adenocarcinoma but not normal breast epithelium
- Proof that the CBFA1 protein in nuclear extracts of breast adenocarcinoma cells is active
- Provisional correlation of CBFA1 expression with the potential for osteotropic metastasis of breast adenocarcinoma

- Demonstration that the mimicry of the osteoblast phenotype by human breast adenocarcinoma cells includes the anomalous production of osteocalcin, a protein normally viewed as an osteoblastic marker
- Isolation from bone metastases of derivative cell lines of the malignant human breast adenocarcinoma model MDA-MB-231 that produce an osteolytic response in the skeletal site
- Identification of cohorts of human genes expressed by human breast adenocarcinoma cells that are significantly upregulated or downregulated by coculture with osteoblast-derived factors

### Reportable Outcomes:

#### Manuscripts, Abstracts, Presentations:

Hecht AC, Adolphson LD, Feinstein MI, Barnes GL, Gerstenfeld LC, George C, Danciu T, Keel S, Skazkina K, Gebhardt MC, and Hauschka PV (1999). Breast Cancer Metastases to Bone Express Increased CBFA1 and Osteocalcin. Proceedings of the Orthopedic Research Society, Abstract. Selected for oral presentation by Dr. Hecht, February 1-4, 1999, Anaheim, CA

Hecht AC, Barnes GL, Yang QL, Adolphson LD, Skazkina K, and Hauschka PV (2000). Mimicry of the osteoblast phenotype by breast adenocarcinoma cells. Abstract. DoD Breast Cancer Research Program Era of Hope Meeting, June 8-12, 2000, Atlanta, GA

Manuscripts on these topics are in preparation and will be submitted following editorial review.

#### Development of Cell Lines:

**M1, M2, and Blast-4** sublines of the MDA-MB-231 malignant human breast adenocarcinoma were recovered, cultured, and passaged during this project from individual bone metastases in nude mice. Subject to further characterization and analysis, these cells will be made available to the scientific community.

### Conclusions:

Our data suggest that 1) the breast adenocarcinoma cell mimics the osteoblast by expressing both osteocalcin and CBFA1, and that 2) regulation of these phenotypic properties may be caused by contact with osteoblast-derived factors in the bone microenvironment. There appears to be increased expression of osteocalcin and CBFA1 in the bone metastasis-derived cell line (M1) when compared to the original MDA-MB-231 tumor line, whereas the normal human breast HMEC cells are negative for both markers. These observations suggest a strong correlation between increasing metastatic potential and the expression of the osteoblast phenotype by the adenocarcinoma cell.

Microcalcification of Breast Lesions: A Possible Consequence of Osteoblastic Mimicry by Adenocarcinoma Cells? An important clinical observation suggesting the existence of a privileged relationship between breast and bone tissue is that deposits of calcium compounds are often seen radiographically and histopathologically among breast carcinoma cells. Ectopic calcifications associated with malignant lesions are formed by hydroxyapatite,  $[\text{Ca}_{10}(\text{PO}_4)_6(\text{OH})_2]$ , the basic mineral found in the skeleton. Microcalcifications are an important indication of early breast carcinoma.<sup>14</sup> In many instances, the detection of microcalcifications on mammography is the unique



sign indicative of the presence of a breast lesion. However, microcalcification deposits in the breast are not restricted to malignant lesions, but can also be associated with benign conditions such as fibroadenoma, secretory diseases and fat necrosis.<sup>15</sup> Ultrastructural analysis of breast cancer associated microcalcifications has revealed crystalline deposits in the cytoplasm of malignant cells, calcification of cytoplasmic organelles, and membrane bound vesicles with hydroxyapatite crystals.<sup>16</sup> These observations have suggested that hydroxyapatite deposition within breast cancer results from an active mechanism rather than the mineralization of cellular debris and necrotic material. Although mammographically detected microcalcifications are frequently the only sign of malignant breast tissue, the mechanism of their deposition has not yet been elucidated. Apparently, breast cancer cells are able to generate a microenvironment that promotes crystallization of calcium and phosphate into bone-like mineral. This observation led investigators to examine breast cancer cells for the expression of bone matrix proteins. Osteonectin, osteopontin and bone sialoprotein have been studied in a series of human breast cancers.<sup>3,15-30</sup> Immunohistochemical evaluation of these three proteins in benign and malignant breast lesions revealed markedly increased expression of BSP/OPN/OSN when compared to benign lesions or normal breast tissue. The detection of OSN and OPN in breast tissues was not surprising due to the ubiquitous presence of these two proteins in many normal and cancerous tissues of the body,<sup>31</sup> however, the expression of BSP was thought to be strictly limited to mineralized connective tissues. BSP was found to be present in numerous breast adenocarcinoma cell lines, both estrogen dependent and independent. In addition, BSP expression was found to be a poor prognostic sign when present in the primary lesion. There was a significantly increased incidence of subsequent bone metastases in patients who expressed higher levels of BSP.<sup>18</sup>

Other matrix proteins may also be important in the metastatic process. Osteocalcin has been implicated, along with BSP, in osteoclast recruitment, activation and attachment. However, there are virtually no reports of non-osteoblastic osteocalcin expression. Our preliminary findings of osteocalcin expression by MDA-MB-231 cells identifies osteocalcin as a potential regulator of osteolytic metastasis.

CBFA1, the Transcriptional Regulator of the Osteoblast Phenotype: The molecular mechanisms involved in the selective interaction of breast cancer cells with the bone matrix are not yet identified. The finding that the minor bone proteins; BSP, OPN and OSN, may play a role in this metastatic process may be further illuminated by the discovery of a novel transcription factor that controls osteoblast differentiation. A transcription factor, CBFA1, was recently shown to control the expression of many osteoblast-specific genes. CBFA1 (core-binding factor), also referred to as PEB2A $\alpha$  (polyoma enhancer-binding protein) and AML-3 (acute myelogenous leukemia) is one of three mammalian genes (i.e. CBFA1, CBFA2, CBFA3) which encode transcription factors whose DNA-binding domains share homology with the *Drosophila* segmentation gene product runt.<sup>32</sup> The alpha subunit of these heterodimeric protein transcription factors binds to DNA via the runt domain when paired with the  $\beta$ -subunit which does not directly interact with the DNA. Recent studies have shown that CBFA1 controls the pathway of differentiation into the osteoblast lineage. The promoter regions of the osteoblast phenotype related genes including OPN, osteocalcin, BSP, and type I collagen contain the core binding sequence.<sup>33-37</sup> Deletion of the CBFA1 gene in mice leads to a total absence of osteoblasts owing to an arrest in their differentiation.<sup>38,39</sup> Komori et al. and Otto et al. independently performed a similar study involving the creation of a CBFA1 knock-out mouse.<sup>37-39</sup> The homozygous CBFA1  $-/-$  mice died soon after delivery, cyanotic, due to inability to breathe. The homozygotes were smaller and had shorter limbs, but all remaining organs were proportionate. The most striking finding was the total lack of bone and retention of the partially calcified cartilaginous skeleton. Membranous bones of the skull and endochondral bone in the

Contains proprietary or unpublished data  
Limit distribution appropriately

skeleton were absent. The tibia contained only calcified cartilage where bone is usually formed at this age. Histology revealed an absence of osteoblasts and smaller sized osteoclasts. Investigators performed northern blot analysis on a number of genes associated with the osteoblast phenotype: osteocalcin, BSP, OPN and alkaline phosphatase. Expression of these genes was greatly diminished. Normal control animals revealed that CBFA1 gene expression was localized during development (via in situ hybridization) to regions destined for bone development. Osteoblast gene expression was examined after BMP-2 stimulation. Both osteocalcin and alkaline phosphatase gene expression was markedly reduced in the mutant calvaria-derived cells. Similarly the genes normally expressed in bone (osteocalcin and osteopontin) were suppressed in CBFA1  $-/-$  mice.

CBFA1 maps to mouse chromosome 17 in mouse and to and to 6p21 in humans<sup>40,41</sup> at the same location as cleidocranial dysplasia (CCD). Otto<sup>39</sup> also noticed that there were abnormalities in heterozygous mice, most prominently: hypoplasia of the clavicle, delayed development of membranous bones, and delayed ossification of cranial bones, causing open anterior and posterior fontanelles, smaller parietal and interparietal cranial bones, and multiple Wormian bones. These features suggested a possible similarity with the human clinical correlate of cleidocranial dysplasia syndrome (CCD). The human CCD syndrome, an autosomal dominant disorder exhibits all the features described above in the CBFA1  $+/-$  mice, plus supernumerary teeth.<sup>42</sup>

CBFA1 was capable of inducing osteoblast related genes in non-osteoblastic tissue. In C3H10t1/2 cells and in skin fibroblasts, transient transfection with a CMV-promoter driven CBFA1 construct up-regulated expression of OPN, osteocalcin, and type I collagen. This essential transcription factor thus controls the lineage specific differentiation of osteoblasts, and may control the breast adenocarcinoma mimicry of the osteoblast phenotype.

We have developed data that point to the role of CBFA1 in controlling the osteoblast phenotype in the metastatic breast adenocarcinoma cell. It is becoming clear that non-osteoblastic cells may be capable of expressing this protein, and it may have different functions in different cell types.<sup>37,43</sup> The unique bone protein osteocalcin was also examined, as CBFA1 directly controls osteocalcin expression. The presence of osteocalcin is indicative of the osteoblast phenotype and this may signify that other key members of the osteoblast program (i.e. PTH/PTHrP receptor, BSP, OPN) are also being expressed by the tumor cells, and that these may also be under the control of CBFA1. The expression of the osteoblast program by the adenocarcinoma cell may allow tumor cells to override the normal osteoblast-osteoclast relationship and to pathologically induce the hormone dependent and cytokine dependent driving of the osteoclast that leads to osteolysis.<sup>46-48</sup>

### **Importance and Implications:**

Bone is the most common site of invasive breast cancer metastasis (over 90% of women dying of breast cancer have bone metastases). The extreme morbidity caused by skeletal metastases (vertebral compression fracture, paraplegia, long bone fracture, severe bone pain) is a major consequence of this disease, causing great personal suffering while consuming an estimated 63% of the total costs of caring for patients with recurrent breast cancer. About 44,300 women died from breast cancer in the U.S. in 1996, and some 184,300 new cases of the disease were diagnosed.<sup>44,45</sup>

We believed that it was essential to focus on defining the limited number mechanisms by which breast adenocarcinoma cells interfere with normal cellular communication in the host skeleton. These mechanisms involve inappropriate ectopic expression of bioactive bone proteins which activate osteoclasts, expression of integrins allowing attachment to the bone ECM, and unregulated

growth factor expression. All clinical protocols which target breast cancer with hormone therapy and cytotoxic chemotherapy must contend with the heterogeneity of the malignant adenocarcinoma phenotype. Diverse mutations and clonal selection of variants evolving in independent sites over the progression of the disease in each patient create an evasive target for chemotherapy. We anticipate that regardless of their detailed phenotypic profile, these malignant cells colonize bone by essentially the same process, and our findings will thus provide a rationale for blocking bone metastasis of breast cancer.

Predictors of future metastasis are of great importance in selecting the high-risk group of *node-negative* breast cancer patients needing careful observation and potentially benefitting from adjuvant therapies. Just as emerging data for microvessel density (angiogenesis) in primary sites correlate well with future recurrence, so may bone protein marker expression eventually predict skeletal metastasis and allow early intervention.

A possible long-term benefit of this Project is the development of molecular approaches to dismantle the pathways by which normal bone cells are coerced into nurturing metastatic cells in the skeletal microenvironment. Importantly, these pathways should be relatively stable targets for therapeutic intervention, in contrast to the highly mutable phenotypes of breast adenocarcinoma cells.

## References

1. Bellahcene A, Merville M-P, Castronovo V (1994). Expression of bone sialoprotein, a bone matrix protein, in human breast cancer. *Cancer Res* 54:2823-6.
2. Bellahcene A, Castronovo V (1995). Increased expression of osteonectin and osteopontin, two bone matrix proteins, in human breast cancer. *Am J Pathol* 146:95-100.
3. Bellahcene A, Kroll M, Liebens F, Castronovo V (1996). Bone sialoprotein expression in primary human breast cancer is associated with bone metastases development. *J Bone Mineral Res* 11:665-70.
4. Hauschka PV, Mavrakos AE, Doleman S, Iafrati M, Klagsbrun M (1986). Growth factors in bone matrix: isolation of multiple types by affinity chromatography on heparin-Sepharose. *J Biol Chem* 261:12665-74.
5. Hauschka PV (1990). Growth factor effects in bone. In Hall BK, ed., *Bone-A Treatise*, v. 1, Telford, pp 103-170.
6. Fisher LW, Hawkins GR, Tuross N, Termine JD (1987). Purification and partial characterization of small proteoglycans I and II, bone sialoproteins I and II, and osteonectin from the mineral compartment of developing human bone. *J Biol Chem* 262:9702-8.
7. Oldberg A, Franzen A, Heinegard D (1988). The primary structure of a cell-binding bone sialoprotein. *J Biol Chem* 263:19430-2.
8. Bianco P, Fisher LW, Young MF, Termine JD, Robey PG (1991). Expression of bone sialoprotein (BSP) in developing human tissues. *Calcif Tissue Int* 49:421-6.
9. Chen J, Shapiro SS, Wrana JL, Reimers S et al. (1991). Localization of bone sialoprotein (BSP) expression to sites of mineralized tissue formation in fetal rat tissues by *in situ* hybridization. *Matrix* 11:133-43.
10. Hauschka PV, Lian JB, Cole DEC, Gundberg CM (1989). Osteocalcin and matrix Gla protein: vitamin K-dependent proteins in bone. *Physiological Rev* 69:990-1047.

11. Ausubel FM, Brent R, Kingston RE, et al. Current Protocols in Molecular Biology. New York: Greene and Wiley, 1989.
12. McCabe LR, Banerjee C, Kundu R, et al. Developmental expression and activities of specific fos and jun proteins are functionally related to osteoblast maturation: Role of fra-2 and jun D during differentiation. *Endocrinology* 1996; 137:4398-4408.
13. Meyers S, Lenny N, Sun W, Hiebert SW. AML-2 is a potential target for transcriptional regulation by the t(8;21) and t(12;21) fusion proteins in acute leukemia. *Oncogene* 1996; 13:303-12.
14. Snyder R, Rosen P. Radiography of breast specimens. *Cancer* 1971; 28:1608.
15. Lanyi M. Microcalcifications in the breast-a blessing or a curse. *Diag Imag Clin Med* 1971; 54:126-145.
16. Ahmed A. Calcification in human breast carcinomas: ultrastructural observations. *J Pathol* 1975; 117:247-251.
17. Bellahcene A, Merville MP, Castronovo V. Expression of bone sialoprotein, a bone matrix protein, in human breast cancer. *Cancer Res* 1994; 54:2823-6.
18. Bellahcene A, Castronovo V. Increased expression of osteonectin and osteopontin, two bone matrix proteins, in human breast cancer. *Am J Pathol* 1995; 146:95-100.
19. Bellahcene A, Menard S, Bufalino R, Moreau L, Castronovo V. Expression of bone sialoprotein in primary human breast cancer is associated with poor survival. *Int J Cancer* 1996; 69:350-3.
20. Bellahcene A, Antoine N, Clausse N, et al. Detection of bone sialoprotein in human breast cancer tissue and cell lines at both protein and messenger ribonucleic acid levels. *Lab Invest* 1996; 75:203-10.
21. Bellahcene A, Castronovo V. Expression of bone matrix proteins in human breast cancer: potential roles in microcalcification formation and in the genesis of bone metastases. *Bull Cancer* 1997; 84:17-24.
22. Gillespie MT, Thomas RJ, Pu ZY, Zhou H, Martin TJ, Findlay DM. Calcitonin receptors, bone sialoprotein and osteopontin are expressed in primary breast cancers. *Int J Cancer* 1997; 73:812-5.
23. Castronovo V, Bellahcene A. Evidence that breast cancer associated microcalcifications are mineralized malignant cells. *Int Jour Onc* 1998; 12:305-308.
24. Bellahcene A, Maloujahnoun N, Fisher LW, et al. Expression of bone sialoprotein in human lung cancer. *Calcif Tissue Int* 1997; 61:183-8.
25. Brown LF, Papadopoulos-Sergiou A, Berse B, et al. Osteopontin expression and distribution in human carcinomas. *Am J Pathol* 1994; 145:610-23.
26. Hirota S, Ito A, Nagoshi J, et al. Expression of bone matrix protein messenger ribonucleic acids in human breast cancers. Possible involvement of osteopontin in development of calcifying foci. *Lab Invest* 1995; 72:64-9.
27. Levedakou EN, Strohmeyer TG, Effert PJ, Liu ET. Expression of the matrix Gla protein in urogenital malignancies. *Int J Cancer* 1992; 52:534-7.
28. Oates AJ, Barraclough R, Rudland PS. The identification of osteopontin as a metastasis-related gene product in a rodent mammary tumour model. *Oncogene* 1996; 13:97-104.
29. van der Pluijm G, Vloedgraven HJ, Ivanov B, et al. Bone sialoprotein peptides are potent inhibitors of breast cancer cell adhesion to bone. *Cancer Res* 1996; 56:1948-55.
30. Tuck AB, O'Malley FP, Singhal H, et al. Osteopontin and p53 expression are associated with tumor progression in a case of synchronous, bilateral, invasive mammary carcinomas. *Arch Pathol Lab Med* 1997; 121:578-84.
31. Brown LF, Berse B, Van de Water L, et al. Expression and distribution of osteopontin in human tissues: widespread association with luminal epithelial surfaces. *Mol Biol Cell* 1992; 3:1169-80.

32. Stewart M, Terry A, Hu M, et al. Proviral insertions induce the expression of bone-specific isoforms of PEBP2alphaA (CBFA1): evidence for a new myc collaborating oncogene. *Proc Natl Acad Sci U S A* 1997; 94:8646-51.
33. Hijiya N, Setoguchi M, Matsuura K, Higuchi Y, Akizuki S, Yamamoto S. Cloning and characterization of the human osteopontin gene and its promoter. *Biochem J* 1994; 303:255-262.
34. Rossert JA, Chen SS, Eberspaecher H, Smith CN, de Crombrughe B. Identification of a minimal sequence of the mouse pro-alpha 1(I) collagen promoter that confers high level osteoblast expression in transgenic mice and binds a protein selectively present in osteoblasts. *Proc Natl Acad Sci* 1996; 93:1027-1031.
35. Towler DA, Bennett CD, Rodan GA. Activity of the rat osteocalcin basal promoter in osteoblastic cells is dependent on homeodomain and CP1 binding motif. *Mol Endocrinol* 1994; 8:614-624.
36. Merriman HL, van Wijnen AJ, Hiebert S, et al. The tissue specific nuclear matrix protein, NMP-2 is a member of the AML/CBF/PEBP2/runt domain transcription factor family; Interactions with the osteocalcin gene promoter. *Biochem* 1995; 34:13125-13132.
37. Ducy P, Zhang R, Geoffroy V, Ridall AL, Karsenty G. Osf2/Cbfa1: a transcriptional activator of osteoblast differentiation [see comments]. *Cell* 1997; 89:747-54.
38. Komori T, Yagi H, Nomura S, et al. Targeted disruption of Cbfa1 results in a complete lack of bone formation owing to maturational arrest of osteoblasts [see comments]. *Cell* 1997; 89:755-64.
39. Otto F, Thornell AP, Crompton T, et al. Cbfa1, a candidate gene for cleidocranial dysplasia syndrome, is essential for osteoblast differentiation and bone development [see comments]. *Cell* 1997; 89:765-71.
40. Levanon D, Negreanu V, Bernstein Y, Bar-Am I, Avivi L, Groner Y. AML1, AML2, and AML3, the human members of the runt domain gene-family: cDNA structure, expression, and chromosomal localization. *Genomics* 1994; 23:425-32.
41. Mundlos S, Otto F, Mundlos C, et al. Mutations involving the transcription factor CBFA1 cause cleidocranial dysplasia [see comments]. *Cell* 1997; 89:773-9.
42. Mundlos S, Mulliken JB, Abramson DL, Warman ML, Knoll JH, BR O. Genetic mapping of cleidocranial dysplasia and evidence of a microdeletion in one family. *Hum Mol Gen* 1995; 4:71-75.
43. Tsuji K, Ito Y, Noda M. Expression of the PEBP2alphaA/AML3/CBFA1 gene is regulated by BMP4/7 heterodimer and its overexpression suppresses type I collagen and osteocalcin gene expression in osteoblastic and nonosteoblastic mesenchymal cells [In Process Citation]. *Bone* 1998; 22:87-92.
44. *Breast Cancer Facts and Figures-1996*. American Cancer Society, Atlanta, 1996.
45. Kosary CL, Ries LAG, Miller BA et al. (1995). *SEER Cancer Statistics Review, 1972-1992: Tables and Graphs*. National Cancer Institute, NIH. Publication 95-2789., Bethesda, MD.
46. Thomas RJ, Guise TA, Yin JJ et al (1999). Breast cancer cells interact with osteoblasts to support osteoclast formation. *Endocrinology* 140:4451-8.
47. Chikatsu N, Takeuchi Y, Tamura Y et al (2000). Interactions between cancer and bone marrow cells induce osteoclast differentiation factor expression and osteoclast-like cell formation in vitro. *Biochem Biophys Res Commun* 267:632-7.
48. Kobayashi K, Takahashi N, Jimi E et al (2000). Tumor necrosis factor alpha stimulates osteoclast differentiation by a mechanism independent of the ODF/RANKL-RANK interaction.

**Appendix****Abstract to 45<sup>th</sup> Orthopedic Research Society, February 1-4, 1999, Anaheim, CA**  
**Accepted for oral presentation.****Breast Cancer Metastases to Bone Express Increased CBFA1 and Osteocalcin.**

Hecht AC, Adolphson LD, Feinstein MI, Barnes GL, Gerstenfeld LC, George C,  
Danciu T, Keel S, Skazkina K, Gebhardt MC, and Hauschka PV.

**Introduction:** Bone Metastases are common in breast cancer. Large autopsy studies estimate their frequency at 85%. Complications of bone metastases include pain, pathologic fracture, loss of mobility and vertebral compression syndromes. The osteotropism of breast cancer metastases remains poorly understood. Several investigators have demonstrated that breast cancer cell lines as well as primary and metastatic tumors synthesize bone proteins such as osteopontin (OPN) and bone sialoprotein (BSP). The transcription factor, CBFA1, regulates the expression of the osteoblast related genes (OC, OPN, BSP). We examined the expression of the osteoblast transcription factor CBFA1 and its target gene osteocalcin (OC) in the MDA-MB 231 cell line, a subclone of this line isolated from a bone metastasis (M1), and archival human specimens of patients with bone metastases from breast cancer primaries. Our central hypothesis was to determine if the expression of the osteoblast phenotype correlated with increased metastatic potential.

**METHODS:** Cell Lines: MDA-MB 231 (ATCC, MD) and Human Mammary Epithelial Cell (HMEC) (Clonetics, CA) were cultured according to recommended guidelines. The M1 subclone was cultured under conditions identical to the MDA-MB 231 cell line. Animals: 7 week old, female Nu/Nu SCID mice (20-25g) were used following institutional guidelines for the use and care of laboratory animals. MDA-MB 231 cells were injected ( $1 \times 10^5$  cells/100ul) through the left second intercostal space into the left ventricle to produce bone metastases. Animals were radiographed bimonthly to detect the presence of lytic lesion. Bones with lesions were fixed in 4% paraformaldehyde and decalcified. Several lesions were isolated and subcultured to produce a breast cancer cell line from a bone metastasis (M1). This subclone was further characterized by immunostaining with cytokeratin 18 (mouse anti-human) to confirm both the human and glandular epithelial origin. Reinjection into mice was performed to confirm its malignancy.

RNA/RT PCR RNA extraction was performed using RNeasy Mini Kit (Quiagen, CA) Primers were synthesized to unique regions in the osteocalcin, CBFA1, and GAPDH human cDNA sequences reported in the Genbank. PCR reaction cycle conditions consisted of denaturation at 93°C for 60s, primer annealing at 61°C for 60s, extension at 72°C for 60s, and a final extension at 72°C for 7 min in a Perkin-Elmer thermocycler (Model 9600). Samples were amplified for 25 cycles. UV photography and densitometry of ethidium bromide stained gels was performed. Semi-quantification was achieved by comparing the intensities of both CBFA1 and osteocalcin to GAPDH.

OC radioimmunoassay. Human OC concentrations in conditioned medium samples (1,3,5,7 days) were measured by radioimmunoassay using monkey anti-human osteocalcin (first antibody) and goat anti-monkey osteocalcin (second antibody). Data are reported as nanograms of OC/ml of medium. Alpha MEM with 10% FCS was used as a control.

Western Blot: Cultures were rinsed with PBS, scraped into buffer and collected by centrifugation. Cells were lysed with hypotonic buffer in the presence of protease inhibitors. Nuclei were collected by centrifugation and resuspended in hypertonic buffer and protease inhibitors. Nuclei were

extracted on ice, insoluble material was cleared by centrifugation and soluble proteins were stored at  $-80^{\circ}\text{C}$ . Nuclear extracts (30ug per lane) were resolved with 8% SDS-PAGE and electroblotted onto nitrocellulose membrane. Membranes were incubated with a 1:100 dilution of antibody in Tris-buffered saline containing 1% BSA. Affinity purified antibodies specific for CBFA1 (gift from Dr. Scott Hiebert, Vanderbilt Cancer Center, Nashville, TN) were used in these studies. Membranes were incubated with secondary antibody followed by streptavidin-HRP chemiluminescence (Pierce, IL).

**Immunohistochemistry:** Monoclonal or polyclonal antibodies to OCN and CBFA1 and appropriate negative controls were employed. The sections were obtained from the distal femur and proximal tibia of mice. Archival human specimens from 10 patients with metastatic breast cancer to appendicular and axial sites were similarly examined. Fixed, paraffin embedded sections from each tumor were immunostained using an indirect avidin-biotin complex method. The appropriate concentration of the primary antibody against CBFA1 and OC were used (1:10 and 1:100).

**Results: RT-PCR.** (Figure 1) Semi-quantitative PCR was performed comparing the expression of CBFA1 and OC between HMEC, MDA-MB 231 cell line, and M1. These results were normalized to GAPDH for each sample. There was a two fold increase in both CBFA1 (375bp) and OCN (294bp) in the M1 clone (lanes 5 and 2) compared to the original MDA-MB 231 cell line (lanes 4 and 1). There was no evidence of CBFA1 in the HMEC (lanes 3 and 6) cell line. PCR product identity was confirmed by producing products of expected size. The CBFA1 amplicon was excised from the gel, purified and underwent automated sequencing to further confirm its identity.

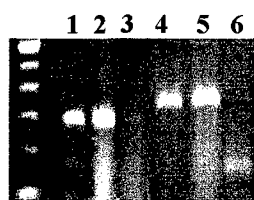


Figure 1

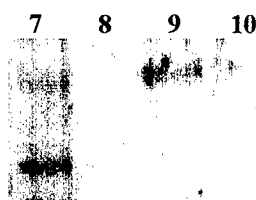


Figure 2

**Western Blot:** (Figure 2) We examined nuclear extracts from the MDA-MB 231 cell line, HMEC line, and M1 subclone, and MG-63 osteosarcoma cell line (positive control) using antibody to CBFA1. CBFA1 was detected in all of these cell lines except the normal breast line HMEC (Lane 8). A 60-65kDa species was present in the MDA-MB 231 line (Lane 10) and M1 subclone (Lane 9). The species was present at a two-fold greater band density in the M1 clone (Lane 9). The MG-63 line (Lane 7) expressed the 60-65 kDa form as well as a 46 kDa form that was not seen in the breast cancer cell lines.

**Radioimmunoassay:** MDA-MB 231 cells produced increasing amounts of osteocalcin. OC concentrations from medium + serum control was measured at 12.8 ng/ml. OC from tumor conditioned medium revealed increasing osteocalcin concentrations. Day 1 produced 14.6 ng/ml; Day 3, 18.4ng/ml, Day 5, 19.7 ng/ml; Day 7, 22.4 ng/ml.

**Immunohistochemistry.** Both osteocalcin and CBFA1 were noted to be positive for immunostaining in both archival human specimens and in experimentally induced animal models of bone metastasis. CBFA1 was seen primarily in the breast cancer metastasis, with minimal staining of the surrounding bone. Both intra and extracellular staining was noted. CBFA1 was noted to be only in the breast cancer metastasis in the archival human specimens with no staining in the surrounding bone. OC was seen in both the breast cancer metastasis and surrounding bone in both the archival human and animal specimens.

**DISCUSSION:** Several studies have found correlations between an increase in osteonectin, OPN and BSP expression with the degree of breast cancer invasiveness. In previous studies, the osteoblast transcription factor CBFA1 has not been found in non-skeletal tissues. CBFA1 plays a pivotal role in osteoblast differentiation and bone formation. This study reveals for the first time that invasive breast cancer is able to synthesize CBFA1 and OC. In addition adenocarcinoma cells isolated from bone expressed a two fold increase in both CBFA1 and OC. The expression of bone related genes may have implications in the pathologic destruction of bone as well as in its osteotropism. OSTEOCALCIN has been shown to increase the recruitment of osteoclasts and promote bone resorption. Paget in 1889 proposed that implantation of a given population of circulating cancer cells at a selected site of the organism is dependent on a suitable environment ("the soil") in which compatible tumor cells ("the seed") could proliferate. The molecular mechanisms underlying this well accepted theory remain poorly understood. The expression of the osteoblast phenotype by the breast cancer cell may give it a survival advantage in the bone microenvironment. In addition, CBFA1 mediates both BSP and OPN expression. These proteins may facilitate the targeting and attachment of circulating tumor cells to areas of ossification. It is tempting to speculate that the expression of CBFA1 may play a key role in explaining the increased osteotropic phenomenon of breast cancer, as well as breast cancer microcalcification,



## **MIMICRY OF THE OSTEOBLAST PHENOTYPE BY BREAST ADENOCARCINOMA CELLS**

**Andrew C. Hecht, George L. Barnes, Quanli Yang, Layla D. Adolphson,  
Katya Skazkina, and Peter V. Hauschka**

Dept. Orthopaedic Surgery, Children's Hospital, Boston, MA 02115

E-mail: [hauschka@hub.tch.harvard.edu](mailto:hauschka@hub.tch.harvard.edu)

Establishment of osteolytic metastases of breast adenocarcinoma in skeletal sites is a complex process wherein tumor cells interfere with normal interactions between osteoblasts and osteoclasts. Mimicry of some osteoblastic functions by malignant adenocarcinoma cells is hypothesized to be central to the biology of the metastatic process in bone.

CBFA1, a transcription factor responsible for normal osteoblast differentiation, is expressed inappropriately by the human breast adenocarcinoma cell line MDA-MB-231, and about 2-fold more strongly by a subline M1, recovered from an osteolytic bone metastasis of MDA-MB-231 in a nu/nu mouse. CBFA1 expression was not detected in normal breast epithelium (HMEC cells). Data were obtained by semiquantitative RT-PCR with specific primers, and by SDS-PAGE immunoblot analysis of nuclear lysates with an antibody that recognizes CBFA1. Electrophoretic mobility shift assay (EMSA) with labeled target oligonucleotide containing the putative binding site for the CBFA1 transcription factor demonstrated band shifting by active CBFA1 protein in nuclear extracts of MB-MDA-231 cells, but not in HMEC. Preincubation with the specific antibody to CBFA1 caused the expected supershifting of the labeled complex to a higher molecular weight.

Expression of CBFA1 by malignant breast adenocarcinoma cells may explain observations of their anomalous expression of "osteoblast-specific" genes (mimicry), because many of these genes have CBFA1 response elements in the 5'-promoter sequences; the osteocalcin gene is a prime example. We analyzed the expression of osteocalcin by SDS-PAGE immunoblotting samples of concentrated 48 hr serum-free conditioned media with specific antibodies that can distinguish between the 6kDa osteocalcin protein sequences of mouse and human. The human breast adenocarcinoma line MCF-7 produced detectable human osteocalcin, while MB-MDA-231 produced marginally detectable levels, and normal HMEC did not appear to express osteocalcin. Co-culture of MCF-7 with a normal mouse osteoblast line (MC3T3-E1) increased the human osteocalcin in the co-culture medium. This effect could be reproduced by addition of conditioned medium from proliferating MC3T3-E1.

We conclude that CBFA1 expression may provide malignant breast adenocarcinoma cells with an advantage in the bone microenvironment, and may explain their observed mimicry of certain aspects of the normal osteoblast. We also infer that the proximity of adenocarcinoma cells to osteoblasts in a skeletal metastasis may allow paracrine effects to enhance mimicry.

The U.S. Army Medical Research and Material Command, under DAMD17-97-1-7114, supported this work.

## **Final Report Data**

### **Bibliography:**

Hecht AC, Adolphson LD, Feinstein MI, Barnes GL, Gerstenfeld LC, George C, Danciu T, Keel S, Skazkina K, Gebhardt MC, and Hauschka PV (1999). Breast Cancer Metastases to Bone Express Increased CBFA1 and Osteocalcin. Proceedings of the Orthopedic Research Society, Abstract. Selected for oral presentation by Dr. Hecht, February 1-4, 1999, Anaheim, CA

Hecht AC, Barnes GL, Yang QL, Adolphson LD, Skazkina K, and Hauschka PV (2000). Mimicry of the osteoblast phenotype by breast adenocarcinoma cells. Abstract. DoD Breast Cancer Research Program Era of Hope Meeting, June 8-12, 2000, Atlanta, GA

Manuscripts on these topics are in preparation and will be submitted following editorial review.

### **List of Personnel Receiving Pay on Research Effort:**

Peter V. Hauschka, Ph.D., P.I.

Andrew C. Hecht, M.D.

Quanli Yang, M.D., Ph.D.

Layla C. Adolphson, B.A.

Katya Skazkina, Ph.D.

Christiane Kuhl, B.S.



DEPARTMENT OF THE ARMY  
US ARMY MEDICAL RESEARCH AND MATERIEL COMMAND  
504 SCOTT STREET  
FORT DETRICK, MARYLAND 21702-5012

*Rec'd  
7/24/2001*

REPLY TO  
ATTENTION OF:

MCMR-RMI-S (70-1y)

MEMORANDUM FOR Administrator, Defense Technical Information  
Center (DTIC-OCA), 8725 John J. Kingman Road, Fort Belvoir,  
VA 22060-6218

SUBJECT: Request Change in Distribution Statement

1. The U.S. Army Medical Research and Materiel Command has reexamined the need for the limitation assigned to technical reports. Request the limited distribution statement for reports on the enclosed list be changed to "Approved for public release; distribution unlimited." These reports should be released to the National Technical Information Service.

2. Point of contact for this request is Ms. Judy Pawlus at DSN 343-7322 or by e-mail at judy.pawlus@det.amedd.army.mil.

FOR THE COMMANDER:

Encl

PHYLLIS M. RINEHART  
Deputy Chief of Staff for  
Information Management